

A Novel Hg²⁺ Selective Ratiometric Fluorescent Chemodosimeter Based on an Intramolecular FRET Mechanism

Gui-Qin Shang · Xia Gao · Mei-Xiu Chen ·
Hong Zheng · Jin-Gou Xu

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Abstract A irreversible Hg²⁺ selective ratiometric fluorescence probe **FR**, a fluorescein fluorophore linked to a rhodamine B hydrazide by a thiourea spacer, was designed and synthesized. The developed probe **FR** exhibited great ratiometric fluorescence enhancement and remarkable yellow-magenta color change toward Hg²⁺ with excellent selectivity in aqueous acetone solution, and the ratiometric fluorescence response to Hg²⁺ was not interfered by other metal cations including Fe³⁺, Co²⁺, Ni²⁺, Cr³⁺, Zn²⁺, Pb²⁺, Cd²⁺, Ca²⁺, Mg²⁺, Ba²⁺ and Mn²⁺. The linear range and the detection limit of this supposed ratiometric fluorescence method for Hg²⁺ were 0.0–10.0×10⁻⁶ and 5×10⁻⁸ M, respectively.

Keywords Fluorescence · Chemosensor · Ratiometric · FRET · Hg²⁺

Introduction

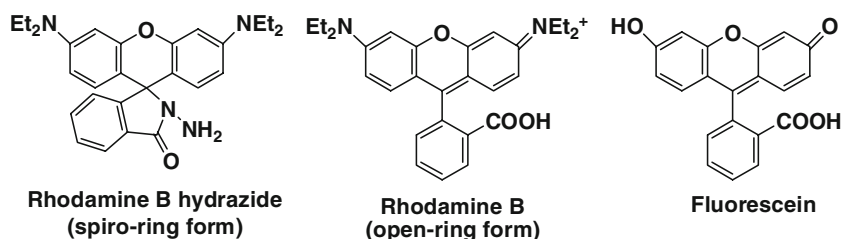
Design and synthesis of fluorescence chemosensors for Hg²⁺ with desirable properties have been of interest to chemists for many years because of the lethal effects of Hg²⁺ on the environment and living organisms [1–4], and a number of chemosensors for Hg²⁺ have been reported [5–19]. Among

these chemosensors, most are direct fluorescence quenching [6–9] or fluorescence enhancement models [10–19]. However, in most practical applications, changes in fluorescence intensity only can also be caused by many other poorly quantified or variable factors such as photobleaching, concentration of probe molecule, the microenvironment around the probe molecule, or the stability of light source. Therefore, there is still an urgent need for feasible chemosensors for the determination of Hg²⁺. Ratiometric method measuring the ratio of fluorescence intensities at two wavelengths provides an alternative approach, which can overcome the drawbacks of intensity-based measurements due to a built-in correction for environmental effects and increase the selectivity, sensitivity and dynamic range of the method [20–25]. However, up to now, only a few ratiometric fluorescence probes for Hg²⁺ [26–28] have been reported in literature. Moreover, the reported sensors were mainly based on the mechanism of intramolecule charge transfer [26–27] or excimer-monomer transfer [28]. To our knowledge, ratiometric fluorescence probe for Hg²⁺ based on fluorescence resonance energy transfer (FRET) has not been reported in literature.

FRET is an interaction between a fluorophore at the electronic excited state (energy donor) and a fluorophore at the ground state (energy acceptor), which leads to the transfer of excitation energy from the donor to the acceptor. Although the efficiency of energy transfer is affected by the distance between the donor and the acceptor and the relative orientation of transition dipoles of both the donor and acceptor, it is also mainly determined by the extent of the spectral overlap between the donor emission and acceptor absorption [29]. Therefore, we thought that it would be possible to fabricate a probe based on the FRET mechanism if a molecule could dramatically generate a suitable fluorescent energy acceptor by the interaction with

G.-Q. Shang · X. Gao · M.-X. Chen · H. Zheng (✉) · J.-G. Xu
Key Laboratory of Analytical Sciences, Ministry of Education,
Department of Chemistry,
College of Chemistry and Chemical Engineering,
Xiamen University,
Xiamen 361005, China
e-mail: hzheng@xmu.edu.cn

Fig. 1 The structures of rhodamine B hydrazide, rhodamine B and fluorescein



target analyte. It was anticipated that a rhodamine B spirolactam derivative would be appropriate candidate because it has two conformations (spirolactam form and ring opened amide form) with distinctly different absorption and fluorescence properties [30]. The ring-opening amide form of rhodamine B derivative has strong absorption at around 560 nm and emits strong fluorescence, whereas the spirolactam form displays absorption only in the ultraviolet region and no fluorescence. Obviously, if a fluorescein derivative is chosen as the energy donor, there is a significant spectral overlap between the emission of fluorescein derivative and the absorption of ring-opening amide form of rhodamine B derivatives (see Figs. 1 and 2).

Based on the above thinking, a ratiometric fluorescence probe **FR** for Hg^{2+} was fabricated, which was composed of fluorescein fluorophore linked to a rhodamine B hydrazide by a thiourea spacer and could function as a dual colorimetric and ratiometric fluorescent reporter for Hg^{2+} . Further experiments showed that **FR** made a feature of good selectivity for Hg^{2+} in water and a red region emission excited at visible wavelength with a ratiometric mode.

Experimental

Apparatus

A Hitachi F-4500 spectrofluorometer (Tokyo, Japan) equipped with a plotter unit and a 1.0 cm quartz cell was used for recording fluorescence spectra and making fluorescence measurements. The absorption spectra were made on a Beckmann DU7400 absorption spectrophotometer (America).

Reagents

All the reagents were used as received from Shanghai Chemicals Group Company except for FITC from Acros Organics. The inorganic salts were of the highest purity available and existed in their nitrates or chlorides. Twice

deionized water was further distilled in the presence of KMnO_4 .

Buffer solution (pH5.0) prepared by mixing 0.2 M 3, 3-dimethylglutaric acid solution and 0.2 M sodium hydroxide solution was used.

The synthesized dye **FR** dissolved in acetone solution to make a 1.0×10^{-3} M stock solution. A 1.0×10^{-3} M standard solution of mercuric chloride was prepared by dissolving 13.6 mg of the reagent in water and diluting up to 50 mL.

The probe **FR** was synthesized as follows (Scheme 1): Rhodamine B hydrazide was synthesized according to the literature [31]. Fluorescein isothiocyanate (FICT, 0.50 g, 1.3 mmol) and rhodamine B hydrazide (0.59 g, 1.3 mmol) were dissolved in 4.0 mL dry dimethylfuran, and the reaction mixture was stirred at room temperature under N_2 atmosphere for 48 h. After removal of the solvent, the residue was purified by flash chromatography with $\text{CHCl}_3/\text{acetone}$ as the eluent to afford **FR** (0.98 g, yield: 89%). The product was further confirmed by the results of ^1H NMR (CD_3OD , 500 MHz), ^{13}C NMR ($\text{DMSO}-d_6$, 400 MHz) and electrospray ionization (ESI) mass spectrometry (MS).

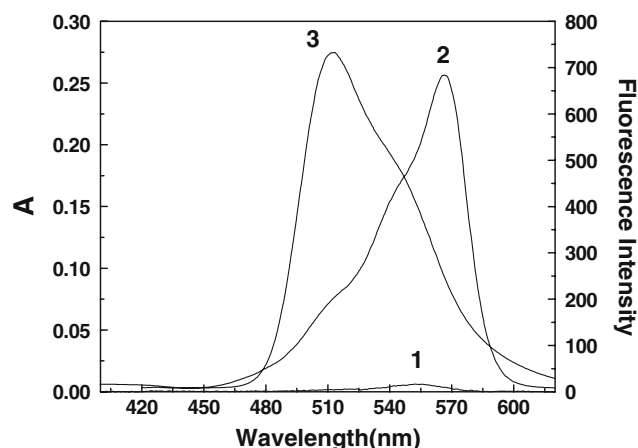
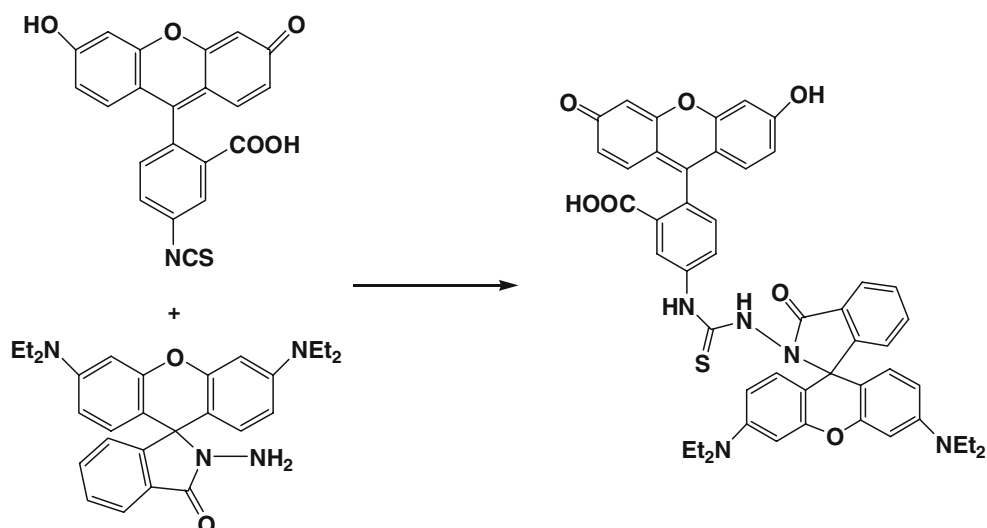


Fig. 2 The absorption spectra of rhodamine B hydrazide and rhodamine B (1 and 2, respectively) and the fluorescence emission spectrum of fluorescein ($\lambda_{\text{ex}}=490$ nm)

Scheme 1 Synthesis of FR



$^1\text{H NMR}$ (CD_3OD , 500 MHz) δ 1.098 (t, $J=8.0$ Hz, 12 H, $4\times\text{NCH}_2\text{CH}_3$), 3.298–3.345 (m, 12 H, including $4\times\text{NCH}_2\text{CH}_3$; and 4 active H: $1\times\text{OH}$, $1\times\text{COOH}$, 2 H in thiourea group, exchangeable with MeOD), 6.328 (d, $J=10.0$ Hz, 2 H), 6.480 (s, 1 H), 6.484 (s, 2 H), 6.502 (d, $J=3.0$ Hz, 2 H), 6.554 (s, 2 H), 6.576 (s, 1 H), 6.654 (d, $J=3.0$, 2 H), 6.954 (d, $J=10.0$ Hz, 1 H), 7.265 (d, $J=9.5$ Hz, 1 H), 7.368 (dd, $J=10.0$, 2.0 Hz, 1 H), 7.666 (s, 1 H), 7.638 (d, $J=9.5$ Hz, 1 H), 7.720 (dt, $J=9.5$, 1.0 Hz, 1 H), 7.996 (d, $J=9.5$ Hz, 1 H)

$^{13}\text{C NMR}$ (DMSO-d_6 , 100 MHz) δ 12.902, 19.019, 30.064, 32.561, 40.900, 44.121, 56.294, 56.500, 63.268, 66.749, 68.979, 83.336, 97.557, 102.763, 104.929, 110.071, 112.975, 123.414, 123.513, 124.729, 126.158,

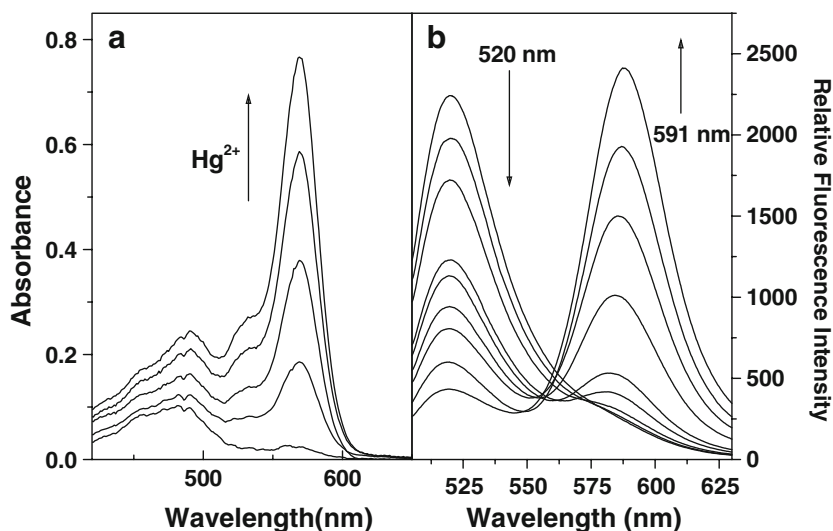
129.157, 130.107, 134.129, 140.947, 148.949, 152.285, 153.995, 159.939, 168.717, 181.700

ESI mass spectrometry, m/z : 846.2 (M^+)

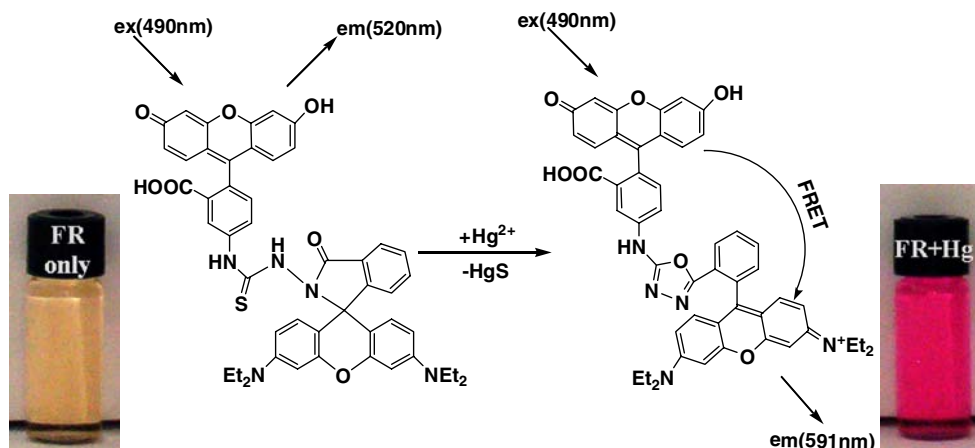
General procedure

Transfer appropriate amounts of HgCl_2 solutions into a series of 10.0 mL volumetric flasks, then add 1.0 mL 3,3-dimethylglutaric acid–NaOH buffer (pH 5.4) and 4.0 mL acetone. Dilute the solution with doubly-distilled water to the mark, then add 100 μL 1.0×10^{-3} M **FR** stock solution with a micropipette and mix thoroughly. After the mixture was incubated at room temperature for 20 min, measure the

Fig. 3 The absorption spectra (a) and fluorescence spectra (b) of FR in the presence of Hg^{2+} . **a** $[\text{FR}]=10.0\ \mu\text{M}$; $[\text{Hg}^{2+}]=0, 8.0, 10.0, 20.0, 30.0\ \mu\text{M}$, respectively. **b** $[\text{FR}]=10.0\ \mu\text{M}$; $[\text{Hg}^{2+}]=0, 1.0, 2.0, 3.0, 5.0, 6.0, 7.0, 10.0\ \mu\text{M}$, respectively. Medium: acetone-water solution (40:60, v/v); pH 5.40



Scheme 2 FRET-based detection mechanism of Hg^{2+}

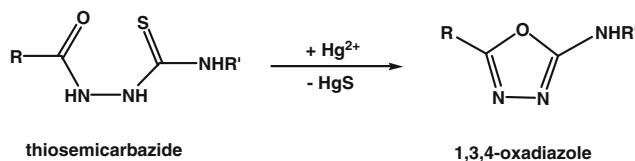


fluorescence ratio of I_{591}/I_{520} with the excitation and emission wavelengths at 490 and 591/520 nm, respectively.

Results and discussions

It can be seen from the curve in Fig. 3a that the free FR ($[\text{Hg}^{2+}] = 0.0 \text{ mol/L}$) showed a maximum absorption wavelength at 490 nm, which exhibited slightly yellow color dominating by the fluorescein chromophore, and no intramolecular FRET phenomenon can be observed in free **FR** because the rhodamine B hydrazide group in **FR** shows only a very small absorption in the wavelength region of the fluorescein emission and this group is nonfluorescent. Therefore, only green fluorescence (520 nm) of fluorescein itself was observed when free **FR** was excited at 490 nm (Fig. 3b, the curve of $[\text{Hg}^{2+}] = 0.0 \text{ mol/L}$).

As reported [32–33], Hg^{2+} promoted the desulfurization reaction of thiosemicarbazide to form 1,3,4-oxadiazole:



Similarly, a thiosemicarbazide group exists in **FR**, therefore, the reaction of Hg^{2+} at the thiosemicarbazide group will force **FR** to form a 1,3,4-oxadiazole group as a new spacer and lead to the release of fluorescent rhodamine B moiety, which triggers an intramolecular FRET. Scheme 2 outlines the reaction mechanism of Hg^{2+} with **FR** based on this design.

The proposed mechanism was further confirmed by the succedent experimental results. Besides the absorption peak

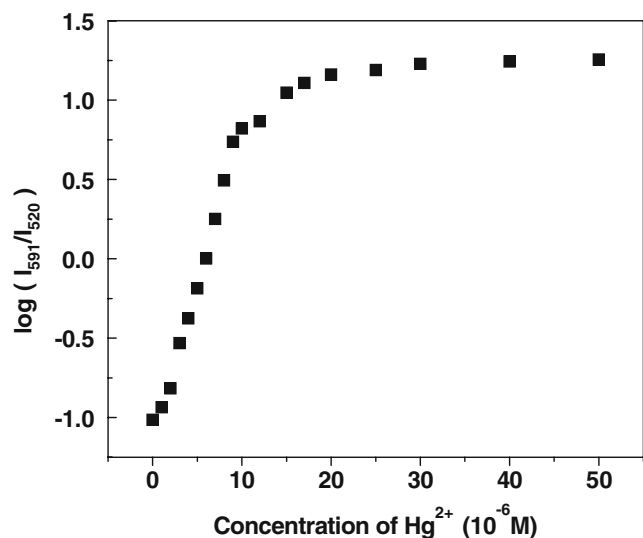
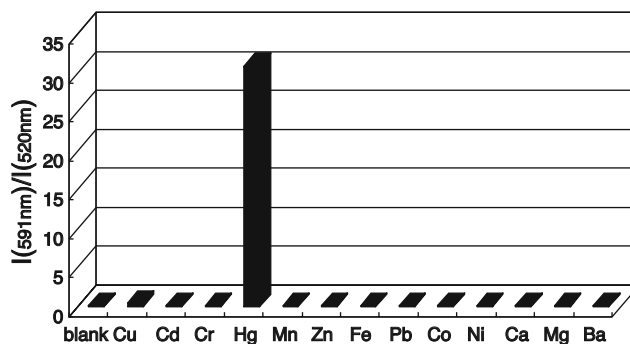


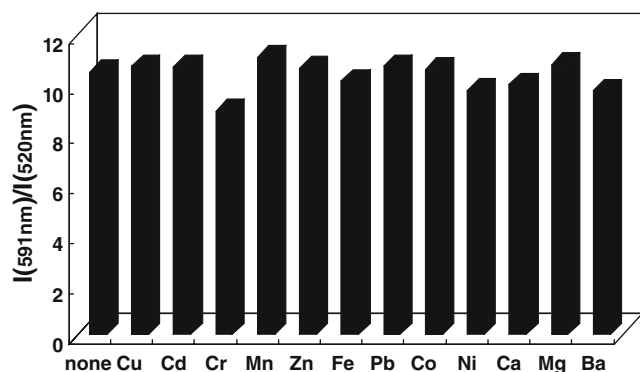
Fig. 4 The titration curves of fluorescent ratio of **FR** vs. Hg^{2+} ionic concentration. $[\text{FR}] = 10 \text{ } \mu\text{M}$; $\lambda_{\text{ex}}/\lambda_{\text{em}} = 490/520, 591 \text{ nm}$

Table 1 Fluorescence ratio of **FR** upon addition of 50.0 μM of various cations



From left to right: no cation (blank), Cu^{2+} , Cd^{2+} , Cr^{3+} , Hg^{2+} , Mn^{2+} , Zn^{2+} , Fe^{3+} , Pb^{2+} , Co^{2+} , Ni^{2+} , Ca^{2+} , Mg^{2+} , Ba^{2+} . $[\text{FR}] = 10.0 \text{ } \mu\text{M}$; Reaction medium: acetone-water solution (40:60, v/v) of pH 5.40

Table 2 Fluorescence intensity of FR (1.0×10^{-5} mol/L) in the presence of mercury ion (1.0×10^{-5} mol/L) and competing ions (5.0×10^{-5} mol/L)



at 490 nm, **FR** showed a new strong absorption peak at 565 nm (Fig. 3a) when the presence of Hg^{2+} , which was attributed to the absorption peak of ring-opening rhodamine B moiety. We have validated the fluorescent product of **FR** after reaction with Hg^{2+} by ESI-MS. The intense peak of m/z 814.1 (M^+) supported the proposed reaction product shown in Scheme 2. Accordingly, this spectral change resulted in the color change from yellow to magenta (Scheme 2), indicating that **FR** can also serve as a highly sensitive “naked-eye” indicator for Hg^{2+} in water.

Meanwhile, great changes in the fluorescence spectrum of **FR** in the presence of Hg^{2+} were also observed (Fig. 3b). The free **FR** displayed a single emission band centered at 520 nm when excited at 490 nm, which was attributed to the emission of fluorescein, whereas, the reaction system of **FR** with Hg^{2+} exhibited dual fluorescence peaks located at 520 and 591 nm, respectively, the latter agreed with the emission of ring-opening rhodamine B moiety. Furthermore, the fluorescence intensity at 520 nm was decreased and the fluorescence intensity at 591 nm was increased at the same time with the increase of Hg^{2+} concentration. Hence, the determination of Hg^{2+} can be performed by measuring the ratio of fluorescence intensities at 591 and 520 nm, respectively. This fact obviously indicates that an intramolecular FRET really exists between the fluorescein moiety and the rhodamine B moiety produced by the action of Hg^{2+} in PR. Figure 4 depicted the plot of the ratiometric fluorescence response of **FR** with the increasing amounts of Hg^{2+} and the presence of 1.0 equiv. of Hg^{2+} gave a ca. 65-fold enhancement in ratiometric value of I_{591}/I_{520} with respect to the metal-free solution.

Table 1 shows the fluorescence responses of **FR** to various background metal ions including some related heavy, transition and main group metal ions. As shown in Table 1, free **FR** (1.0×10^{-5} M) exhibits a rather low value of fluorescence ratio (signed as the blank), upon addition of 5.0 equiv. of tested background metal ions, the fluorescence

ratio is nearly not affected, while the addition of Hg^{2+} results in a large ratio value, indicating the high fluorimetric selectivity for Hg^{2+} .

Furthermore, interferences from various coexistent metal cations for the determination of $10.0 \mu\text{M}$ Hg^{2+} ion were also investigated. The fluorescence ratio value of **FR** in the presence of $10.0 \mu\text{M}$ Hg^{2+} ion was almost unaffected (relative error $\leq \pm 10\%$) by the addition of 5.0 equiv of competing metal ions except for Cr^{3+} caused an approximately -15% relative error (Table 2).

Conclusion

In summary, we have developed a new fluorescence probe, **FR**, for Hg^{2+} based on an intramolecular FRET with a high selectivity. The color of this probe changes from yellow to magenta when reacted with Hg^{2+} , which makes it available to detect Hg^{2+} either by ratiometric fluorimetry or by rapid “naked eye” detection. Moreover, the ratiometric fluorescence detection for Hg^{2+} provides a built-in correction for environmental effects, which is in favor of serving as a practical probe for rapid and accurate determination of mercuric ion in environmental systems. Furthermore, the successful fabrication of the proposed probe provides an alternative concept to design ratiometric fluorescence probes by utilizing two fluorophores satisfying the requirements of intramolecular FRET.

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References

- Desvergne JP, Czarnik AW (1997) Chemosensors of Ion and Molecule Recognition. Kluwer, Dordrecht
- Czarnik AW (ed) (1992) In: Fluorescent Chemosensors for Ion and Molecule Recognition. American Chemical Society, Washington, D.C.
- Hashem EY (2002) Spectrophotometric studies on the simultaneous determination of cadmium and mercury with 4-(2-pyridylazo)-resoreinol. Spectrochim Acta Part A 58:1401–1410
- Chatterjee S, Pillai A, Gupta VK (2002) Spectrophotometric determination of mercury in environmental sample and fungicides based on its complex with o-carboxy phenyl diazaminophenyl p-azobenzene. Talanta 57:461–465
- Ines O, Otto SW (1997) Optical sensors for determination of heavy metal ions. Microchimica Acta 126(3/4):177–192
- Rurack K, Resch-Genger U, Bricks JL, Spieles M (2000) Cation-triggered ‘switching on’ of the red/near infra-red (NIR) fluorescence of rigid fluorophore-spacer-receptor ionophores. Chem Commun 2:2103–2104
- Talanova GG, Elkarim NSA, Talanov VS (1999) A calixarene-based Fluorogenic reagent for selective mercury(II) recognition. Anal Chem 71:3106–3109

8. Kim JS, Gil Choi M, Song KC, Tai No K, Ahn S, Chang SK (2007) Ratiometric determination of Hg^{2+} ions based on simple molecular motifs of pyrene and dioxaoctanediamide. *Org Lett* 9:1129–1132
9. Zhang XB, Guo CCh, Li ZZ, Shen GL, Yu RQ (2002) An optical fiber chemical sensor for mercury ions based on a porphyrin dimer. *Anal Chem* 74(4):821–825
10. Torsten M, Christian I, Gregor L, Ingo K, Otto SW (2003) Cross-reactive metal ion sensor array in a micro titer plate format. *Anal Chem* 75(17):4389–4396
11. Chan WH, Yang RH, Wang KM (2001) Development of a mercury ion-selective optical sensor based on fluorescence quenching of 5,10,15,20-tetraphenylporphyrin. *Anal Chim Acta* 444(2):261–269
12. Wu FY, Zhao YQ, Ji ZJ, Wu YM (2007) A Highly Sensitive and Selective Fluorescent Chemodosimeter for Hg^{2+} in Neutral Aqueous Solution. *J Fluoresc* 17:460–465
13. Yang XF, Li Y, Bai Q (2007) A highly selective and sensitive fluorescein-based chemodosimeter for Hg^{2+} ions in aqueous media. *Anal Chim Acta* 584:95–100
14. Andrew AV, Ramaier N (1998) Optical fibre reflectance sensors for the detection of heavy metal ions based on immobilised Br-PADAP. *Sens Actuators B Chem* B51(1–3):368–376
15. Ivana M, Otto SW (1997) Fluorescence-based sensor membrane for mercury (II) detection. *Sens Actuators B Chem* B39(1–3):246–251
16. Sasaki DY, Padilla BE (1998) Dithioamide metal ion receptors on fluorescent lipid bilayers for the selective optical detection of mercuric ion. *Chem Commun* 18:1581–1582
17. Nolan EM, Lippard SJ (2003) A “turn-on” fluorescent sensor for the selective detection of mercuric ion in aqueous media. *J Am Chem Soc* 125:14270–14271
18. Guo X, Qian X, Jia L (2004) A highly selective and sensitive fluorescent chemosensor for Hg^{2+} in neutral buffer aqueous solution. *J Am Chem Soc* 126:2272–2273
19. Yang YK, Yook KJ, Tae J (2005) A rhodamine-based fluorescent and colorimetric chemodosimeter for the rapid detection of Hg^{2+} ions in aqueous media. *J Am Chem Soc* 127:16760–16761
20. Kubo Y, Yamamoto M, Ikeda M, Takeuchi M, Shinkai S, Yamaguchi S, Tamao K (2003) A colorimetric and ratiometric fluorescent chemosensor with three Emission changes: fluoride ion sensing by a triarylborane–porphyrin conjugate. *Angew Chem Int Ed* 42:2036–2040
21. Raker J, Glass TE (2002) Selectivity via cooperative interactions: detection of dicarboxylates in water by a pinwheel chemosensor. *J Org Chem* 67:6113–6116
22. Fu H, Loo BH, Xiao D, Xie R, Ji X, Yao J, Zhang B, Zhang L (2002) Multiple emissions from 1,3-diphenyl-5-pyrenyl-2-pyrazoline nanoparticles: evolution from molecular to nanoscale to bulk materials. *Angew Chem Int Ed* 41:962–965
23. Mohr GJ, Klimant I, Spichiger UE, Wolfbeis OS (2001) Fluoro reactands and dual luminophore referencing: a technique to optically measure amines. *Anal Chem* 73:1053–1056
24. Mello JV, Finney NS (2001) Dual-signaling fluorescent chemosensors based on conformational restriction and induced charge transfer. *Angew Chem Int Ed* 40:1536–1538
25. Takakusa H, Kikuchi K, Urano Y, Sakamoto S, Yamaguchi K, Nagano T (2002) Design and synthesis of an enzyme-cleavable sensor molecule for phosphodiesterase activity based on fluorescence resonance energy transfer. *J Am Chem Soc* 124:1653–1657
26. Coskun A, Deniz Yilmaz MU, Akkaya E (2007) Bis(2-pyridyl)-substituted boratriazaindacene as an NIR-emitting chemosensor for $\text{Hg}(\text{II})$. *Org Lett* 9:607–609
27. Liu B, Tian H (2005) A selective fluorescent ratiometric chemodosimeter for mercury ion. *Chem Commun* 25:3156–3158
28. Moon SY, Youn NJ, Park SM, Chang SK (2005) Diametrically disubstituted cyclam derivative having Hg^{2+} -selective fluoroionophoric behaviors. *J Org Chem* 70:2394–2397
29. Lakowicz JR (1999) Principles of Fluorescence Spectroscopy, 2nd edn. Plenum, New York
30. Nguyen T, Francis MB (2003) A practical synthetic route to functionalized rhodamine dyes. *Org Lett* 5:3245–3248
31. Dujols V, Ford F, Czarnik AW (1997) A long-wavelength fluorescent chemodosimeter selective for $\text{Cu}(\text{II})$ ion in water. *J Am Chem Soc* 119:7386–7387
32. Wang X, Li Z, Wei B, Yang J (2002) Synthesis of 2-(4-methoxyphenyloxy-acetylamido)-5-aryloxymethyl-1,3,4-oxadiazoles under microwave irradiation. *Synth Commun* 32:1097–1103
33. Zou X, Jin G (2001) Synthesis of pyridazinone-substituted 1,3,4-thiadiazoles-1,3,4-oxadiazoles and 1,2,4-triazoles. *J Heterocycl Chem* 38:993–996